

**Amendments to the Claims:**

Please amend the claims as follows:

1. (Currently amended) Process for generating and detecting recombinant DNA sequences in prokaryotes comprising the steps of:

- (a) generating a first prokaryotic cell containing ~~[[a]]an extrachromosomal~~ recipient DNA molecule, which comprises a first DNA sequence to be recombined and which can autonomously replicate in the prokaryotic cell, and ~~[[a]]an extrachromosomal~~ donor DNA molecule, which comprises a second DNA sequence to be recombined and at least a first marker sequence encoding a gene product and which cannot autonomously replicate in the prokaryotic cell,
- (b) cultivating the first prokaryotic cell under selective conditions, ~~which force the formation of a co-integrate or hybrid molecule between the recipient and donor DNA molecules and the recombination of the two DNA sequences to be recombined and which only allow the growth and/or propagation of the cell if the gene product of the first marker sequence is expressed, and~~
- (c) isolating a second prokaryotic cell grown and/or propagated under selective conditions and containing a hybrid DNA molecule with the at least first marker sequence and a first and a second recombined DNA sequence~~[[s]]~~ due to recombination between the first and the second DNA sequences~~[[.]]~~,

wherein the prokaryotic cell is transiently or permanently deficient in the mismatch repair system.

2. (Currently amended) Process according to claim 1, wherein the donor DNA molecule and the recipient DNA molecules are different linear or circular DNA structures, in particular different plasmids or bacteriophages~~[[.]]~~.

3. (Currently amended) Process according to claim 1, wherein the recipient DNA molecule is a plasmid, which can replicate in *Escherichia coli*, donor DNA molecule does not have an origin of replication.
4. (Currently amended) Process according to claim 3, wherein the recipient DNA molecule is the *E. coli* plasmid pACYC184 or the *E. coli* plasmid pMIX100 or a derivative thereof, 1, wherein the donor DNA molecule has a non-functional origin of replication.
5. (Currently amended) Process according to claim 1 where the donor DNA molecule does not have an origin of replication, wherein the donor DNA molecule is a *Bacillus subtilis* plasmid, which cannot replicate in *E. coli*.
6. (Currently amended) Process according to claim 1 where the donor DNA molecule has a non-functional origin of replication 5, wherein the donor DNA molecule is the *B. subtilis* plasmid pMIX91 comprising the *spec<sup>R</sup>* marker and the *phleo<sup>R</sup>* marker or the *B. subtilis* plasmid pMIX101 comprising the *tc<sup>R</sup>* marker.
7. (Currently amended) Process according to claim 6, wherein the donor DNA molecule and/or its origin of replication are derived from a prokaryotic species other than the prokaryotic species, in cells of which the donor DNA molecule is introduced 1, wherein the first marker sequence of the donor DNA structure is selected from the group consisting of a nutritional marker, an antibiotic resistance marker and a sequence encoding a subunit of an enzyme.
8. (Currently amended) Process according to claim 7, wherein the donor DNA molecule is a *Bacillus subtilis* plasmid, which cannot replicate in *E. coli* gene product of the first marker sequence confers resistance to an antibiotic to a cell which is sensitive to that antibiotic.
9. (Currently amended) Process according to claim 7, where the donor DNA molecule is the *B. subtilis* plasmid pMIX91 comprising the *spec<sup>R</sup>* marker and the *phleo<sup>R</sup>* marker or the *B. subtilis* plasmid pMIX101 comprising the *tc<sup>R</sup>* marker wherein the first marker sequence is *spec<sup>R</sup>*, the gene product of which confers to a cell

resistance to spectinomycin, or *phleo*<sup>R</sup>, the gene product of which confers to a cell resistance to phleomycin, or *tc*<sup>R</sup>, the gene product of which confers to a cell resistance to tetracycline.

10. (Currently amended) Process according to claim 6, wherein the function of the origin of replication of the donor DNA is impaired by a mutation. Process for generating and detecting recombinant DNA sequences in prokaryotes comprising the steps of:

- d) generating a first prokaryotic cell containing an extrachromosomal recipient DNA molecule, which comprises a first DNA sequence to be recombined and which can autonomously replicate in the prokaryotic cell, and an extrachromosomal donor DNA molecule, which comprises a second DNA sequence to be recombined and at least a first marker sequence encoding a gene product and which cannot autonomously replicate in the prokaryotic cell,
- e) cultivating the first prokaryotic cell under selective conditions, which force the formation of a co-integrate or hybrid molecule between the recipient and donor DNA molecules and the recombination of the two DNA sequences to be recombined and which only allow the growth and/or propagation of the cell if the gene product of the first marker sequence is expressed, and
- f) isolating a second prokaryotic cell grown and/or propagated under selective conditions and containing a hybrid DNA molecule with the at least first marker sequence and a first and a second recombined DNA sequence due to recombination between the first and the second DNA sequences,

wherein the donor DNA molecule is the *B. subtilis* plasmid pMIX91 comprising the *spec*<sup>R</sup> marker and the *phleo*<sup>R</sup> marker or the *B. subtilis* plasmid pMIX101 comprising the *tc*<sup>R</sup> marker.

11. (Currently amended) Process according to claim 4 wherein the first marker sequence of the donor DNA structure is selected from the group consisting of a nutritional marker, an antibiotic-resistance marker and a sequence encoding a

subunit of an enzyme, 10, wherein the recipient DNA molecule is a linear or circular DNA structure, in particular a plasmid or a bacteriophage.

12. (Currently amended) Process according to claim 4, wherein the gene product of the first marker sequence confers resistance to an antibiotic to a cell which is sensitive to that antibiotic 10, wherein the prokaryotic cell has a functional mismatch repair system.

13. (Currently amended) Process according to claim 11, wherein the first marker sequence is *spec<sup>R</sup>* the gene product of which confers to a cell resistance to spectinomycin or *phleo<sup>R</sup>* the gene product of which confers to a cell resistance to phleomycin or *tsr<sup>R</sup>* the gene product of which confers to a cell resistance to tetracycline 10, wherein the prokaryotic cell is transiently or permanently deficient in the mismatch repair system.

14. (Currently amended) Process according to claim 1, wherein the donor DNA molecule contains a second marker sequence, recipient DNA molecule is a plasmid, which can replicate in *Escherichia coli*.

15. (Currently amended) Process according to claim 4, wherein the recipient DNA molecule contains a third marker sequence and optionally a fourth marker sequence 14, wherein the recipient DNA molecule is the *E. coli* plasmid pACYC184 or the *E. coli* plasmid pMIX100 or a derivative thereof.

16. (Currently amended) Process according to claim 14, wherein the second, third and fourth marker sequences are protein-coding or non-coding sequences selected from the group consisting of nutritional markers, pigment markers, antibiotic resistance markers, antibiotic sensitivity markers, restriction enzymes sites, primer recognition sites and sequences encoding a subunit of an enzyme 1, wherein the donor DNA molecule and/or its origin of replication are derived from a prokaryotic species other than the prokaryotic species in cells of which the donor DNA molecule is introduced.

17. (Currently amended) Process according to claim 16, wherein the gene products of the third and fourth marker sequences of the recipient DNA molecule confer resistance to an antibiotic to a cell which is sensitive to that antibiotic 1, wherein the function of the origin of replication of the donor DNA is impaired by a mutation.

18. (Currently amended) Process according to claim 17, wherein the gene product of the third marker sequence confers to a cell resistance to tetracycline 1, wherein the donor DNA molecule contains a second marker sequence.

19. (Currently amended) Process according to claim 17, wherein the gene product of the fourth marker sequence confers to a cell resistance to chloramphenicol 1, wherein the recipient DNA molecule contains a third marker sequence and optionally a fourth marker sequence.

20. (Currently amended) Process according to claim 1, wherein the first and the second DNA sequences to be recombined diverge by at least two nucleotides 18, wherein the second, third and fourth marker sequences are protein-coding or non-coding sequences selected from the group consisting of nutritional markers, pigment markers, antibiotic resistance markers, antibiotic sensitivity markers, restriction enzymes sites, primer recognition sites and sequences encoding a subunit of an enzyme.

21. (Currently amended) Process according to claim 1, wherein the first and the second DNA sequences to be recombined are naturally occurring sequences 20, wherein the gene products of the third and fourth marker sequences of the recipient DNA molecule confer resistance to an antibiotic to a cell which is sensitive to that antibiotic.

22. (Currently amended) Process according to claim 21, wherein the first and/or the second DNA sequences to be recombined are derived from viruses, bacteria, plants, animals and/or human beings, gene product of the third marker sequence confers to a cell resistance to tetracycline.

23. (Currently amended) Process according to claim 1, wherein the first and/or the second DNA sequences to be recombined are artificial sequences 21, wherein the gene product of the fourth marker sequence confers to a cell resistance to chloramphenicol.
24. (Currently amended) Process according to claim 1, wherein each of the first and the second DNA sequences to be recombined comprises one or more protein-coding sequences and/or one or more non-coding sequences diverge by at least two nucleotides.
25. (Currently amended) Process according to claim 1, wherein the first prokaryotic cell is generated by simultaneously or sequentially introducing the recipient DNA molecule and the donor DNA molecule into a prokaryotic cell and the second DNA sequences to be recombined are naturally occurring sequences.
26. (Currently amended) Process according to claim 25, wherein the recipient and donor DNA molecules are introduced into the prokaryotic cell via transformation, conjugation, transduction, sexduction and/or electroporation first and/or the second DNA sequences to be recombined are derived from viruses, bacteria, plants, animals and/or human beings.
27. (Currently amended) Process according to claim 10, wherein the first prokaryotic cell is cultivated in the presence of at least one antibiotic to which the gene product of the first marker sequence confers resistance 1, wherein the first and/or the second DNA sequences to be recombined are artificial sequences.
28. (Currently amended) Process according to claim 27, wherein the first prokaryotic cell is additionally cultivated in the presence of a second, a third and/or a fourth antibiotic to which the gene products of the second marker sequence, the third marker and the fourth marker sequence, respectively, confer resistance 1, wherein each of the first and the second DNA sequences to be recombined comprises one or more protein-coding sequences and/or one or more non-coding sequences.

29. (Currently amended) Process according to claim 1, wherein the prokaryotic cell is a cell of an archaeabacterium or an eubacterium, first prokaryotic cell is generated by simultaneously or sequentially introducing the recipient DNA molecule and the donor DNA molecule into a prokaryotic cell.

30. (Currently amended) Process according to claim 29, wherein the eubacterium is a gram-negative bacterium, a gram-positive bacterium or a cyanobacterium, recipient and donor DNA molecules are introduced into the prokaryotic cell via transformation, conjugation, transduction, sexduction and/or electroporation.

31. (Currently amended) Process according to claim 30, wherein the gram-negative bacterium is *Escherichia coli* 1, wherein the first prokaryotic cell is cultivated in the presence of at least one antibiotic to which the gene product of the first marker sequence confers resistance.

32. (Currently amended) Process according to claim 4, wherein the prokaryotic cell has a functional mismatch repair system 31, wherein the first prokaryotic cell is additionally cultivated in the presence of a second, a third and/or a fourth antibiotic to which the gene products of the second marker sequence, the third marker sequence, and the fourth marker sequence, respectively, confer resistance.

33. (Currently amended) Process according to claim 1, wherein the prokaryotic cell is transiently or permanently deficient in the mismatch repair system, a cell of an archaeabacterium or an eubacterium.

34. (Currently amended) Process according to claim 33, wherein the transient or permanent deficiency of the mismatch repair system is due to a mutation, a deletion, and/or an inducible expression or repression of one or more genes involved in the mismatch repair system, a treatment with an agent that saturates the mismatch repair system and/or a treatment with an agent that globally knocks out the mismatch repair eubacterium is a gram-negative bacterium, a gram-positive bacterium or a cyanobacterium.

35. (Currently amended) Process according to claim 33, wherein the prokaryotic cell has a mutated *mutS* gene and/or mutated *mutL* gene 34, wherein the gram-negative bacterium is *Escherichia coli*.

36. (Currently amended) Process according to claim 1, wherein the first and the second recombined DNA sequences contained in the hybrid DNA molecule of the second prokaryotic cell are selected and/or isolated and/or analysed transient or permanent deficiency of the mismatch repair system is due to a mutation, a deletion and/or an inducible expression or repression of one or more genes involved in the mismatch repair system, a treatment with an agent that saturates the mismatch repair system and/or a treatment with an agent that globally knocks out the mismatch repair system.

37. (Currently amended) Process according to claim 36, wherein the first and the second recombined DNA sequences are isolated by restriction enzyme cleavage 1, wherein the prokaryotic cell has a mutated *mutS* gene and/or mutated *mutL* gene.

38. (Currently amended) Process according to claim 36, wherein the first and the second recombined DNA sequences are amplified by PCR 1, wherein the first and the second recombined DNA sequences contained in the hybrid DNA molecule of the second prokaryotic cell are selected and/or isolated and/or analysed.

39. (Currently amended) Process according to claim 36, wherein the isolated first and second recombined DNA sequences are inserted into a donor DNA molecule and a recipient DNA molecule, respectively, and subjected another round of recombination 38, wherein the first and the second recombined DNA sequences are isolated by restriction enzyme cleavage.

40. (Currently amended) *Bacillus subtilis* plasmid pMIX81 which comprises the *spec<sup>R</sup>* marker and the *phleo<sup>R</sup>* marker and the restriction sites *ScaI*, *PpuMI* and *EcoO109I* for inserting a foreign DNA sequence. Process according to claim 38, wherein the first and the second recombined DNA sequences are amplified by PCR.

41. (Currently amended) Bacillus subtilis plasmid pMIX101 which comprises the tc<sup>R</sup>-marker sequence and the restriction sites XbaI and PstI for inserting a foreign DNA sequence. Process according to claim 38, wherein the isolated first and second recombined DNA sequences are inserted into a donor DNA molecule and a recipient DNA molecule, respectively, and subjected to another round of recombination.

42. (Currently amended) Use of the B. subtilis plasmids pMIX91 or pX101 as donor-DNA molecules in a process according to claim 1, for generating and/or detecting recombinant DNA sequences in a prokaryotic host cell, preferably in an E. coli cell. Bacillus subtilis plasmid pMIX91, which comprises the spec<sup>R</sup> marker and the phleo<sup>R</sup> marker and the restriction sites SalI, PpuMI and EcoO109I for inserting a foreign DNA sequence.

43. (Currently amended) Use of the E. coli-plasmids pACYC184 or pMIX100 or a derivative thereof as recipient-DNA molecule in a process according to claim 1, for generating and/or detecting recombinant DNA sequences in a prokaryotic host cell, preferably in an E. coli cell. Bacillus subtilis plasmid pMIX101 which comprises the tc<sup>R</sup> marker sequence and the restriction sites XbaI and PstI for inserting a foreign DNA sequence.

44. (Cancelled)

45. (Cancelled)

46. (Currently amended) A process for producing a hybrid gene and/or a protein encoded by a hybrid gene in a prokaryotic cell, wherein a process according to claim 4 is carried out and the hybrid gene and/or the protein encoded by the hybrid gene is produced in the prokaryotic cell and the hybrid gene and/or the encoded protein is selected in the prokaryotic cell and/or isolated therefrom after expression. Kit comprising at least a first container which comprises cells of the E. coli strain AB1157 or the E. coli strain MXP1 or the E. coli strain DHB10, a second container which comprises cells of the E. coli strain AB1157 containing plasmid pACYC184 or cells of the E. coli strain DHB10 containing plasmid pMIX100 and a third container

comprising cells of the *B. subtilis* strain DSM17158 containing plasmid pMIX91 or cells of the *B. subtilis* strain DSM17159 containing pMIX101.

47. (Currently amended) Hybrid gene obtainable by a process according to claim

46. Kit comprising at least a first container which comprises cells of the *E. coli* strain AB1157 or the *E. coli* strain MXP1 or the *E. coli* strain DHB10, a second container comprising DNA of plasmid pACYC184 or plasmid pMIX100 and a third container comprising DNA of plasmid pMIX91 or plasmid pMIX101.

48. (Currently amended) Protein, which is encoded by a hybrid gene according to claim 47 and which is obtainable by a process according to claim 46. A process for producing a hybrid gene and/or a protein encoded by a hybrid gene in a prokaryotic cell, wherein a process according to claim 1 is carried out and the hybrid gene and/or the protein encoded by the hybrid gene is produced in the prokaryotic cell and the hybrid gene and/or the encoded protein is selected in the prokaryotic cell and/or isolated therefrom after expression.